

Formation Mechanism of *p*-Methylacetophenone from Citral via a *tert*-Alkoxy Radical Intermediate

TOSHIO UENO,^{*,†} HIDEKI MASUDA,[†] AND CHI-TANG HO[‡]

Material Research and Development Laboratories, Ogawa & Company, Ltd., 15-7 Chidori, Urayasushi, Chiba 279-0032, Japan, and Department of Food Science, Rutgers University, 65 Dudley Road, New Brunswick, New Jersey 08901-8520

The aim of this study was to clarify the formation mechanism of a potent off-odorant, *p*-methylacetophenone, from citral under acidic aqueous conditions. An acidic aqueous solution (pH 3.0) containing 10 mg/L of citral was stored at 40 °C for 2 weeks. Among the compounds detected in the stored citral solution, 4-(2-hydroxy-2-propyl)benzaldehyde was identified for the first time as a degradation product from citral. The formation of *p*-methylacetophenone and 4-(2-hydroxy-2-propyl)benzaldehyde behaved the same when antioxidants were added to the citral solution. In addition, both compounds were formed by the Fe²⁺-induced decomposition of 8-hydroperoxy-*p*-cymene, another compound identified in the stored citral solution. These results suggested that both *p*-methylacetophenone and 4-(2-hydroxy-2-propyl)benzaldehyde can be formed via the same radical intermediate [*p*-CH₃C₆H₄C(CH₃)₂O•] that can be derived from the O–O bond homolysis of 8-hydroperoxy-*p*-cymene. On the other hand, the degradation of 8-hydroperoxy-*p*-cymene without Fe²⁺ under acidic aqueous conditions did not yield *p*-methylacetophenone and 4-(2-hydroxy-2-propyl)benzaldehyde, but the degradation of citral without Fe²⁺ did. Therefore, other than the decomposition of 8-hydroperoxy-*p*-cymene, a mechanism to generate the *tert*-alkoxy radical intermediate was proposed for the formation of *p*-methylacetophenone and 4-(2-hydroxy-2-propyl)benzaldehyde in the citral solution.

KEYWORDS: Citral; degradation; off-odors; *p*-methylacetophenone; formation mechanism; *tert*-alkoxy radical; 4-(2-hydroxy-2-propyl)benzaldehyde

INTRODUCTION

The stability of citral (a mixture of neral and geranial) under acidic aqueous conditions is a critical issue in the field of flavor chemistry. Under such conditions, citral easily degrades by a series of cyclization and oxidation reactions to form a variety of degradation products (1–8). Consequently, not only is the fresh lemon-like odor of citral lost, but undesirable off-odors develop. Among the degradation products from citral, *p*-cresol and *p*-methylacetophenone were reported to be the most potent off-odorants (6, 7). The contribution of these two compounds to the off-odors formed in the acidic solution of citral was also confirmed in our experiments (9).

Despite the importance as off-odorants derived from citral, the formation mechanisms of *p*-cresol and *p*-methylacetophenone have not yet been fully clarified. **Scheme 1** shows the previously proposed formation pathways of the oxidation products including *p*-cresol and *p*-methylacetophenone from citral under acidic aqueous conditions. It has been established that citral in acidic solutions undergoes cyclization reactions leading to the formation of *p*-menthadien-8-ols (1–5, 8).

p-Cymen-8-ol and its dehydration product, α ,*p*-dimethylstyrene, have been proposed to be formed from *p*-menthadien-8-ols by their oxidation with dissolved oxygen (5) or disproportionation reactions (4) producing α -terpineol as the reduction product. *p*-Methylacetophenone was then suggested to be formed by the oxidation of α ,*p*-dimethylstyrene (6). In our previous studies, on the other hand, 8-hydroperoxy-*p*-cymene was detected in the stored acidic solution of citral, and it was demonstrated that this hydroperoxide decomposes to *p*-cresol, *p*-cymen-8-ol, and α ,*p*-dimethylstyrene, and not to *p*-methylacetophenone (9) under the same acidic conditions as the citral solution. The formation mechanism of *p*-methylacetophenone from citral under our experimental conditions thus remains a question.

Therefore, the aim of this study was to clarify the formation mechanism of *p*-methylacetophenone from citral under acidic aqueous conditions.

MATERIALS AND METHODS

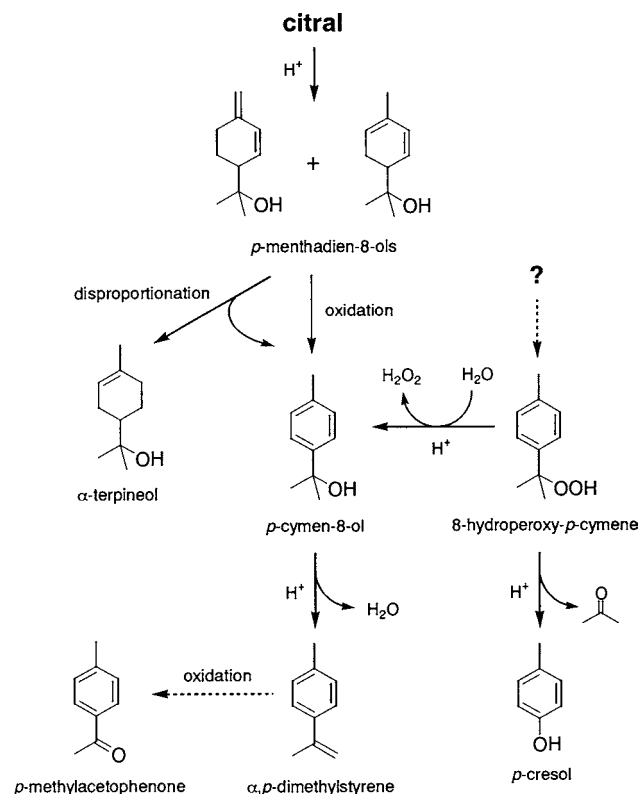
Chemicals. Citral was purchased from Polarome International (Jersey City, NJ). *p*-Cymen-8-ol and α ,*p*-dimethylstyrene were purchased from Sigma-Aldrich Japan (Tokyo, Japan). (–)-Epicatechin (EC), (–)-epicatechin gallate (ECg), (–)-epigallocatechin (EGC), (–)-epigallocatechin gallate (EGCg), and (+)-catechin were purchased from Funakoshi (Tokyo, Japan). *p*-Cymene and quercitrin were purchased from Wako Pure Chemical Industries (Osaka, Japan). *p*-Cresol, *p*-

* Author to whom correspondence should be addressed (fax +81-47-305-1423; e-mail ueno.toshio@ogawa.net).

[†] Ogawa & Co., Ltd.

[‡] Rutgers University.

Scheme 1. Previously Proposed Formation Pathways of *p*-Cresol and *p*-Methylacetophenone and Other Oxidation Products from Citral under Acidic Aqueous Conditions (4–6, 9)



methylacetophenone, and ascorbic acid were purchased from Nacalai Tesque (Kyoto, Japan).

Synthesis. 4-(2-Hydroxy-2-propyl)benzaldehyde was synthesized from *p*-bromobenzaldehyde dimethylacetal (Sigma-Aldrich Japan) according to the method described by Creary and Wang (10). A yield of 13 mg (31%) of 4-(2-hydroxy-2-propyl)benzaldehyde was obtained: 1H MMR ($CDCl_3$) δ 10.00 (s, 1 H), 7.76 (AA'BB' aromatic quartet, 4H), 2.16 (br s, 1H), 1.61 (s, 6H) [agreed with the literature (10)]; MS (EI), m/z (relative intensity) 164 (2), 150 (10), 149 (100), 145 (5), 133 (3), 121 (4), 115 (5), 107 (6), 105 (5), 91 (7), 77 (14), 74 (4), 65 (3), 59 (6), 51 (10), 43 (94).

8-Hydroperoxy-*p*-cymene was synthesized as follows: A solution of 150 mg of *p*-cymen-8-ol in 5 mL of ethanol was added dropwise to the stirred mixture of 150 mL of 30% hydrogen peroxide and 15 mL of 2.5% (w/v) sulfuric acid. After stirring for 1 h at room temperature, the reaction mixture was extracted with 250 mL of dichloromethane, washed with water (250 mL \times 2), dried over sodium sulfate, and concentrated in vacuo to \sim 1 mL. The crude product was purified by thin-layer chromatography on silica gel plates using a mixture of ethyl acetate and hexane (1:5, v/v) as the eluent to give 8-hydroperoxy-*p*-cymene (38 mg, yield 23%). The MS (EI) and 1H NMR spectra were consistent with the literature values (11, 12).

Model Reactions. A sample solution containing 10 mg/L of citral in an acidic buffer (0.1 M citric acid/0.2 M sodium hydrogen phosphate, pH 3.0) with or without 60 mg/L of an added antioxidant (Table 2) was prepared. One hundred milliliters of the prepared solution was transferred to a 100-mL glass bottle, and the sample bottle was sealed with a Teflon liner and a screw cap. The sample was stored in a dark incubator at 40 °C for 2 weeks. For studies of the metal-induced decomposition of 8-hydroperoxy-*p*-cymene, 1 mL of 5×10^{-2} M ferrous sulfate heptahydrate in water was added to 100 mL of the sample solution containing 2 mg/L of 8-hydroperoxy-*p*-cymene in the acidic buffer. The color of the solution immediately turned yellow. The sample bottle was shaken and then left at room temperature for 30 min.

Preparation of Analytical Samples. The degradation products of citral or those of 8-hydroperoxy-*p*-cymene were extracted with dichloromethane (30 mL \times 2). One milliliter of 0.01% (w/v) *n*-pentadecane

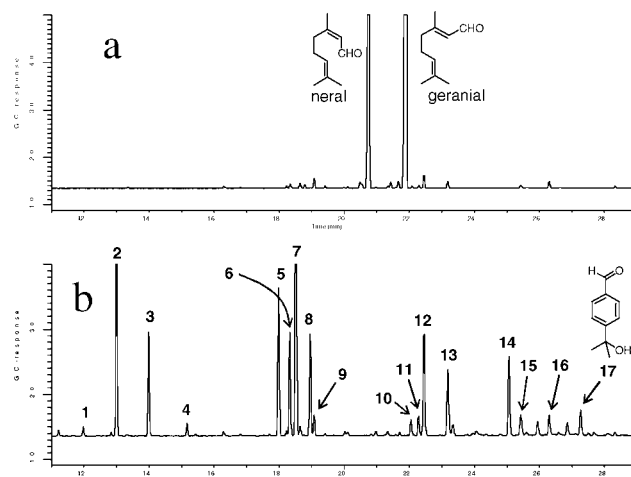


Figure 1. Gas chromatographic analysis of the acidic buffer solution (pH 3.0) of citral (10 mg/L) (a) before and (b) after storage at 40 °C for 2 weeks. Numbers correspond to those in Table 1.

in dichloromethane was added to the extract as the internal standard. The extract was dried over sodium sulfate, concentrated in vacuo to \sim 5 mL, and further concentrated under a stream of nitrogen to \sim 200 μ L.

Gas Chromatography (GC). An Agilent 6890 N gas chromatograph equipped with a flame ionization detector (FID) and a DB-1 fused silica capillary column (60 m \times 0.25 mm i.d.; film thickness of 0.25 μ m; J&W Scientific) was used. To minimize the decomposition of 8-hydroperoxy-*p*-cymene, an injector temperature of 100 °C was used. The other operating conditions were as follows: detector temperature, 250 °C; nitrogen carrier gas flow rate, 1 mL/min; oven temperature program, 80 °C, raised at 3 °C/min to 210 °C (15 min); 1 μ L of sample was injected using a split ratio of 1:50. The amounts of the citral degradation products were estimated by computing the areas versus that of the internal standard (*n*-pentadecane). The response factors of all compounds to the FID were assumed to be the same.

Gas Chromatography–Mass Spectrometry (GC-MS). A Hewlett-Packard 5890 series II gas chromatograph equipped with an HP-5972 mass selective detector and a DB-1 fused silica capillary column (60 m \times 0.25 mm i.d.; film thickness of 0.25 μ m; J&W Scientific) was used. The operating conditions were as follows: injector temperature, 250 °C; helium carrier gas flow rate, 1 mL/min; oven temperature program, 60 °C, raised at 3 °C/min to 210 °C (40 min); 1 μ L of sample was injected, using a split ratio of 1:50; ionization voltage, 70 eV; ion source temperature, 140 °C.

Nuclear Magnetic Resonance (NMR) Spectroscopy. 1H NMR spectra were recorded at 400 MHz on a Bruker Avance 400 spectrometer (Bruker, Tsukuba, Japan) in $CDCl_3$ with TMS as internal standard ($=$ 0 ppm).

RESULTS AND DISCUSSION

Degradation of Citral under Acidic Aqueous Conditions. During storage under acidic aqueous conditions, citral (neral and geranial) was completely degraded and almost totally converted to its cyclization products 1–17 (Figure 1 and Table 1), among which 4-(2-hydroxy-2-propyl)benzaldehyde (17) was identified for the first time as a degradation product from citral. The structural estimation based on the MS spectra and subsequent synthesis according to the published method (10) led to the identification of 17. The synthetic sample exhibited an odor character similar to that of benzaldehyde. Because of the weak odor, compound 17 was assumed not to contribute to the off-odors of the stored citral solution.

It has been accepted that compounds 1, 2, 5, 8, 12, 13, 15, and 16 are formed from citral by a series of acid-catalyzed reactions (1–5, 8). In agreement with most published data,

Table 1. Degradation Products Detected in the Acidic Buffer Solution (pH 3.0) of Citral (10 mg/L) Stored at 40 °C for 2 Weeks

no. ^a	compound	RI ^b	ID ^c method
1	2,3-dehydro-1,8-cineole	985	B
2	<i>p</i> -cymene	1017	A
3	<i>p</i> -cresol	1044	A
4	α , <i>p</i> -dimethylstyrene	1077	A
5	<i>p</i> -mentha-1,5-dien-8-ol	1150	B
6	<i>p</i> -methylacetophenone	1158	A
7	<i>p</i> -cymen-8-ol	1163	A
8	<i>p</i> -mentha-1(7),2-dien-8-ol	1174	B
9	α -terpineol	1177	A
10	(2 <i>R</i> ,5 <i>R</i>)-2-formylmethyl-2-methyl-5-(1-hydroxy-1-methylethyl)tetrahydrofuran	1249	C
11	(2 <i>S</i> ,5 <i>R</i>)-2-formylmethyl-2-methyl-5-(1-hydroxy-1-methylethyl)tetrahydrofuran	1254	C
12	<i>trans-p</i> -menth-2-ene-1,8-diol	1259	B
13	<i>cis-p</i> -menth-2-ene-1,8-diol	1276	B
14	8-hydroperoxy- <i>p</i> -cymene	1321	A
15	<i>trans-p</i> -menth-1-ene-3,8-diol	1330	B
16	<i>cis-p</i> -menth-1-ene-3,8-diol	1351	B
17	4-(2-hydroxy-2-propyl)benzaldehyde	1365	A

^a Numbers correspond to those in Figure 1. ^b Retention index on DB-1 (60 m).

^c Identification methods: A, mass spectrum and retention index agree with those of authentic compounds; B and C, compounds were tentatively identified on the basis of the following criteria: (B) mass spectrum agrees with that of the Wiley mass spectral database (Agilent Technologies, 2000) and RI agrees with literature value (5) or (C) mass spectrum agrees with literature spectrum (13).

p-menthadien-8-ols (5 and 8) and *p*-menth-2-ene-1,8-diols (12 and 13) were among the major acid-catalyzed reaction products

in the citral solution. Unlike the studies conducted at room temperature (1, 2, 5), a dehydration product, *p*-cymene (2), was also one of the major products in our experiments, probably due to the relatively high temperature (40 °C) of our experimental conditions.

Oxidation reactions as well as the acid-catalyzed reactions would be involved in the formation of compounds 3, 4, 6, 7, 9–11, 14, and 17. Compounds 10 and 11 were reported to be formed via the direct epoxidation of the 6,7-double bond of citral (13, 14). Kimura et al., on the other hand, demonstrated the formation of α ,*p*-dimethylstyrene (4), *p*-cymen-8-ol (7), and α -terpineol (9) using isolated *p*-menthadien-8-ols 5 and 8, respectively, under acidic aqueous conditions (4). It was unclear, however, whether the other oxidation products, that is, *p*-cresol (3), *p*-methylacetophenone (6), 8-hydroperoxy-*p*-cymene (14), and the newly found 17, would be formed via the major acid-catalyzed degradation products of citral, such as 5 and 8, or would be directly formed from citral.

Figure 2 shows the concentration changes of citral (neral and geranial, Figure 2a), its major acid-catalyzed degradation products (Figure 2b), and the oxidation products (Figure 2c,d) in the citral solution during storage. Neral and geranial almost totally degraded during the first 3 days of storage (Figure 2a). The concentration of acid-catalyzed degradation products 5, 8, 12, and 13 rapidly increased during this period (Figure 2b) and then started decreasing with almost the entire loss of citral, whereas the concentration of their dehydration product 2 kept increasing during the storage. On the other hand, the concentration of the oxidation products 3, 6, 7, and 14 (Figure 2c) and 4 and 17 (Figure 2d) also kept increasing after almost the entire loss of citral, although compound 14 started to decrease after 7

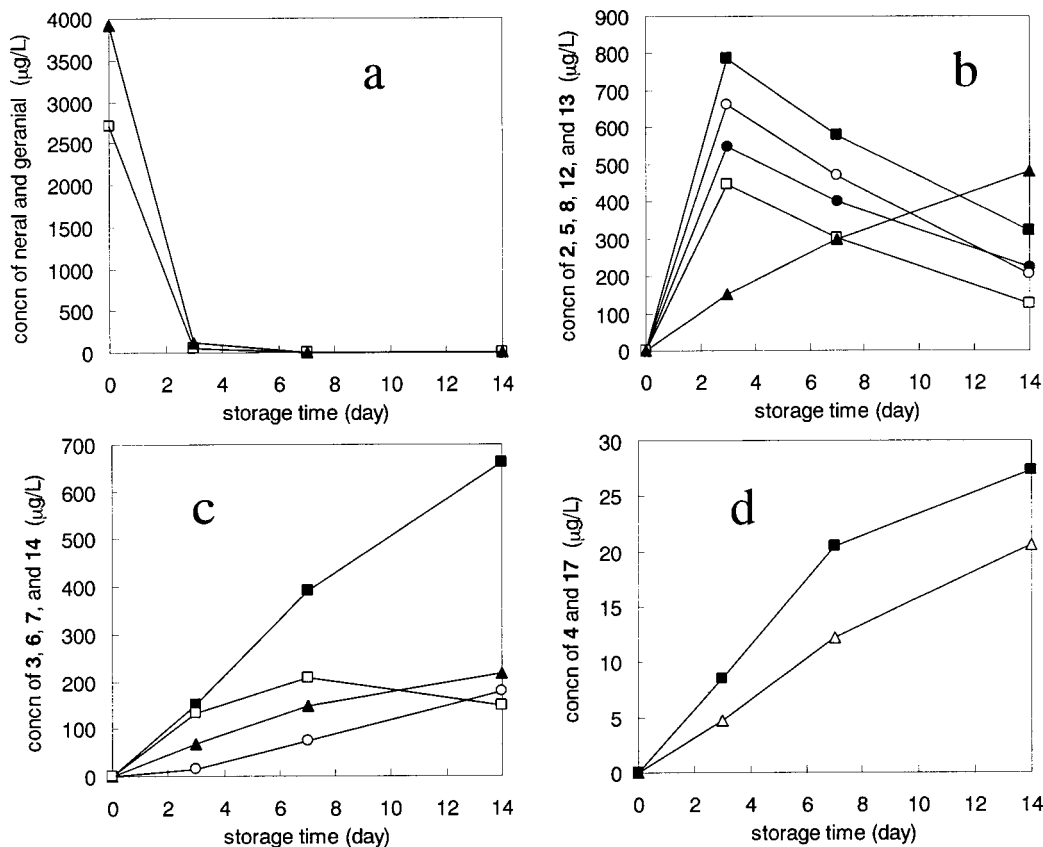


Figure 2. Concentration changes of (a) neral (□) and geranial (▲), of (b) acid-catalyzed degradation products 2 (▲), 5 (■), 8 (●), 12 (○), and 13 (□), of (c) oxidation products 3 (○), 6 (▲), 7 (■), and 14 (□), and of (d) oxidation products 4 (△) and 17 (■) during the storage of citral (10 mg/L) in an acidic buffer solution (pH 3.0) at 40 °C. Each point is the mean value of three experiments. Coefficient of variation for each point was <10%.

Table 2. Effects of Antioxidant Addition on the Formation of the Oxidation Products (**3**, **4**, **6**, **7**, **14**, and **17**) from Citral under Acidic Aqueous Conditions

antioxidant ^b	concn of compound ^a ($\mu\text{g/L}$)					
	3	4	6	7	14	17
control	197	24	209	677	171	29
(+)-catechin	170 ^c	68	23 ^c	1589	134	tr ^d
(-)-epicatechin (EC)	344	42	45	1018	278	3.2
(-)-epigallocatechin (EGC)	299	65	29	1402	263	tr
(-)-epicatechin gallate (ECg)	592	28	21	805	618	tr
(-)-epigallocatechin gallate (EGCg)	556	30	28 ^c	835	565	tr
quercitrin	514	22	106	624	625	11
ascorbic acid	132	23	137	654	152	19

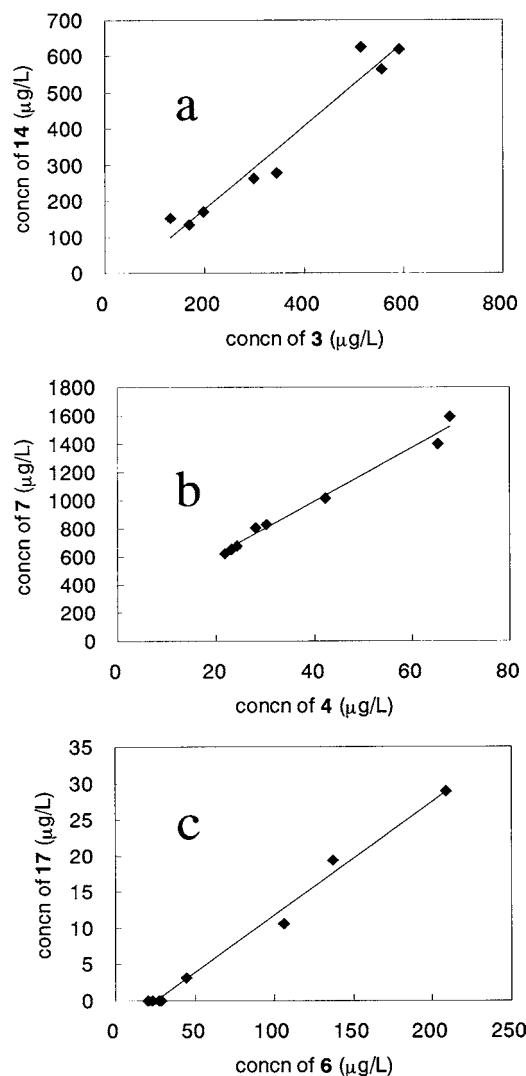
^a Concentration was determined after the storage of the citral solution at 40 °C for 2 weeks. Each value is the mean of five experiments. Coefficient of variation for each value was <10% unless otherwise specified. ^b Sixty milligrams per liter of an antioxidant was added to an acidic buffer solution (0.1 M citric acid/0.2 M sodium hydrogen phosphate, pH 3.0) containing 10 mg/L of citral. ^c Coefficients of variation ranged from 10 to 20%. ^d Trace.

days of storage, probably due to its degradation to **3**, **4**, and **7** under the acidic aqueous conditions (9). These results indicate that the oxidation products **3**, **6**, **14**, and **17** as well as **4** and **7** would be formed not directly from citral but via the major acid-catalyzed cyclization products from citral.

Antioxidant Additions and the Resulting Formation Correlations among Oxidation Products from Citral. The use of antioxidants to inhibit the formation of off-odorants from citral in acidic solutions has already been attempted (4, 5, 9). It was reported (5) that the addition of isoascorbic acid to an acidic solution containing citral strongly inhibited the formation of α ,*p*-dimethylstyrene (**4**) and *p*-cymen-8-ol (**7**). We previously reported that catechins from green tea showed strong inhibitory effects on the formation of *p*-methylacetophenone (**6**) from citral under acidic aqueous conditions (9). In the present study, information regarding such antioxidant additions was examined to investigate the formation pathways of oxidation products from citral.

Table 2 shows the effects of the added antioxidants (60 mg/L) including the catechins, quercitrin, and ascorbic acid on the formation of **3**, **4**, **6**, **7**, **14**, and **17** from citral under acidic aqueous conditions. Among these results, three pairs of oxidation products from citral, that is, **3** and **14**, **4** and **7**, and **6** and **17**, showed linear relationships between their concentrations (**Figure 3**). This means that the formation of the two oxidation products in each pair was influenced in the same manner by this series of antioxidant additions, reflecting the formation pathways of the oxidation products from citral. Therefore, it is not surprising that good correlations were obtained between the concentrations of **3** and **14** (correlation coefficient, $R = 0.975$) and between those of **4** and **7** ($R = 0.993$). These can be explained by their reported formation pathways, in which **3** is formed from **14** and **4** is formed from **7** by acid-catalyzed reactions (**Scheme 1**). It is more interesting to see a good correlation between the concentrations of **6** and **17** ($R = 0.995$, a trace amount of **17** was assumed to be zero), because this might suggest a possible connection between the potent off-odorant **6** and newly found product **17** during their formation from citral.

Fe²⁺-Induced Decomposition of 8-Hydroperoxy-*p*-cymene (14**).** It is well-known that hydroperoxides can undergo homolytic O–O bond cleavage by accepting one electron from reducing metal ions such as ferrous ion (Fe²⁺) to produce alkoxy radicals (15). In the case of the hydroperoxide **14**, the homolytic O–O bond cleavage would produce the corresponding *tert*-alkoxy radical [*p*-CH₃C₆H₄C(CH₃)₂O•], which can undergo

**Figure 3.** Linear relationships between the concentrations of (a) compounds **3** and **14**, (b) **4** and **7**, and (c) **6** and **17** in the citral solution stored with antioxidants (data from **Table 2**).**Table 3.** Quantitative Data for the Fe²⁺-Induced Decomposition of 8-Hydroperoxy-*p*-cymene (**14**) under Acidic Aqueous Conditions

sample ^a	concn of compound ($\mu\text{g/L}$)			
	6	7	14	17
before addition of Fe ²⁺	9	32	2085	nd ^b
after addition of Fe ²⁺	1378	273	nd	278

^a FeSO₄ (5×10^{-4} M) was added to the acidic buffer solution (pH 3.0) of **14** (2 mg/L). ^b Not detected.

further decomposition with the elimination of a methyl radical, that is, β -fragmentation (16), leading to the formation of *p*-methylacetophenone (**6**). To confirm the formation of **6** and other products resulting from the O–O bond homolysis of **14**, we conducted the Fe²⁺-induced decomposition of **14** under acidic aqueous conditions. As shown in **Figure 4**, hydroperoxide **14** immediately degraded to **6** along with **7** and **17** by the addition of a large excess of ferrous sulfate (FeSO₄). The quantitative data for this experiment are given in **Table 3**.

As shown in **Scheme 2**, all of the products **6**, **7**, and **17** were interpreted to be formed via the *tert*-alkoxy radical **18** generated by the reduction of **14** with Fe²⁺. *p*-Cymen-8-ol (**7**) could be formed by the further reduction of the alkoxy radical **18** by

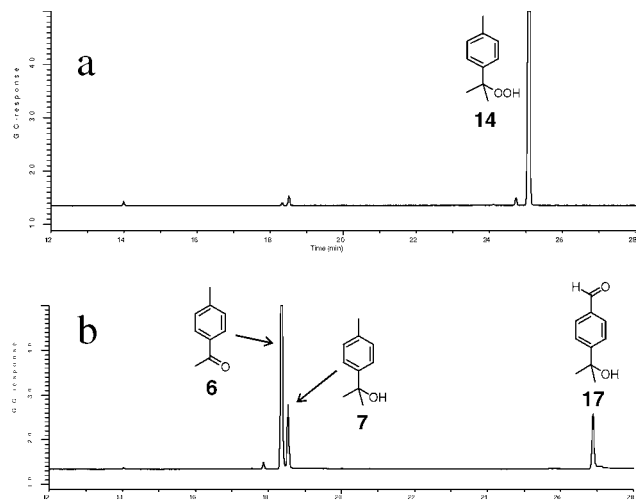
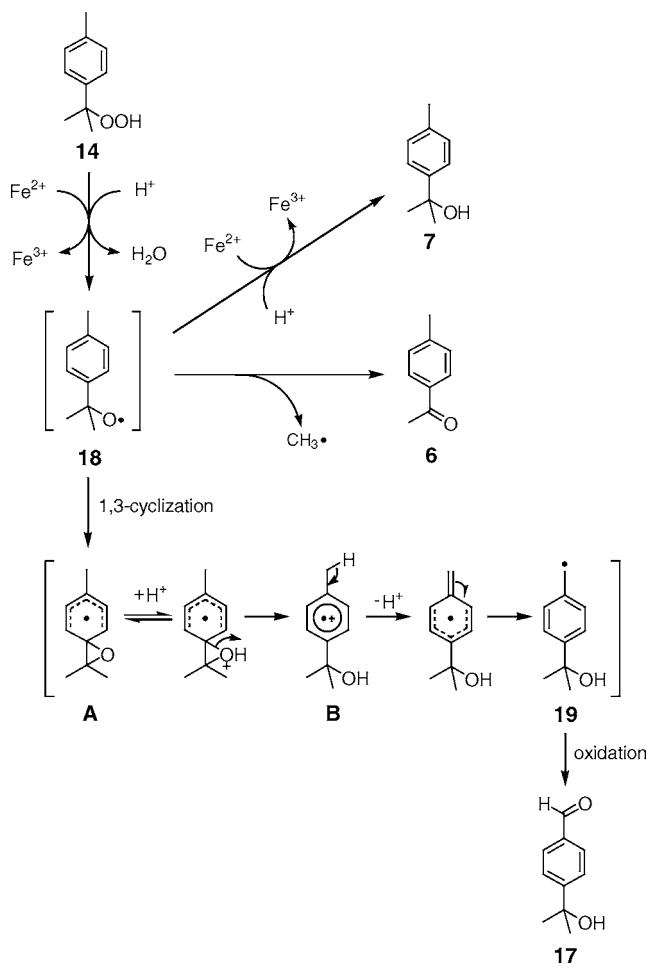


Figure 4. Gas chromatographic analysis of the acidic buffer solution (pH 3.0) of hydroperoxide **14** (2 mg/L) (a) before and (b) after the addition of Fe^{2+} (5×10^{-4} M).

Scheme 2. Possible Mechanism for the Fe^{2+} -Induced Decomposition of Hydroperoxide **14** under Acidic Aqueous Conditions



Fe^{2+} (15). Regarding the formation of **17** from **18**, the transformation of **18** to a benzylic radical **19** was postulated to be involved. A possible mechanism for this transformation is as follows. The alkoxy radical **18** could undergo 1,3-cyclization to form a cyclohexadienyl radical **A** (17), which might convert via the acid-catalyzed ring opening to an aryl radical cation **B**. The

Table 4. Concentration of Degradation Products Obtained from the Acidic Buffer Solutions (pH 3.0) of 8-Hydroperoxy-*p*-cymene (**14**) (2 mg/L), Citral (10 mg/L), and Citral (10 mg/L) with **14** (2 mg/L) Stored at 40 °C for 2 Weeks

sample	concn of compound ^a ($\mu\text{g/L}$)					
	3	4	6	7	14	17
14	849 ^b	25	17	685	127	nd ^c
citral	183	23 ^d	224	678	153	27
citral with 14	1017	48	317	1410	324	41

^a Concentration was determined after the storage of the sample solutions at 40 °C for 2 weeks. Each value is the mean of three experiments. Coefficient of variation for each value was <10% unless otherwise specified. ^b Coefficient of variation was 13%. ^c Not detected. ^d Coefficient of variation was 23%.

aryl radical cation **B** could typically undergo deprotonation at the benzylic position to form the benzylic radical **19** (18, 19).

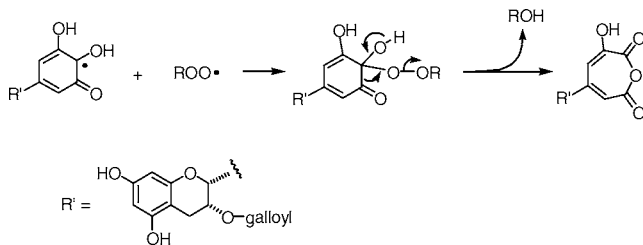
Another possibility for the formation of **19** might be H abstraction at the benzylic position of **7** with free radicals. However, we did not detect *p*-formylacetophenone, an oxidation product that could be obtained from H abstraction at the benzylic position of **6**. This observation suggested that H abstraction from **7** would also not occur. Therefore, we assumed that **19** would be formed from **18** as mentioned above.

Degradation of Hydroperoxide 14 under Acidic Aqueous Conditions without Fe^{2+} . In contrast to the Fe^{2+} -induced decomposition, there was little or no production of **6** and **17** during the storage of **14** under acidic aqueous conditions in the absence of Fe^{2+} (9). This result indicated that **14** without Fe^{2+} rarely undergoes the O–O bond homolysis under our experimental conditions. In this paper, we also report the degradation of **14** in the acidic aqueous solution of citral. **Table 4** shows quantitative data for the degradation products from **14** (2 mg/L), citral (10 mg/L), and citral (10 mg/L) with added **14** (2 mg/L) stored under acidic aqueous conditions. With the addition of **14** to the citral solution, the formation of **3** and **7** (and **4**) increased almost as much as expected from the data of the degradation of **14** by itself. The formation of **6** and **17** in the citral solution was also increased with the addition of **14**, but the increased amounts were much lower than those of **3** and **7**. These results suggested that the degradation of **14** would not be, at least, the main pathway for the formation of **6** and **17** in the citral solution.

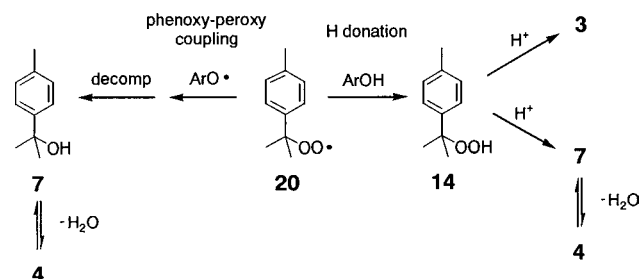
Formation Mechanisms of *p*-Methylacetophenone (6) and Compound 17 from Citral. On the basis of the presented data, we propose the formation mechanisms of **6** and **17** in the citral solution as shown in **Scheme 3**. Our results suggested that both **6** and **17** could be formed via the *tert*-alkoxy radical **18**, yet the decomposition of **14** would not be the main pathway for the formation of **6** and **17** in the citral solution. Therefore, it seems reasonable to assume that, other than the O–O bond homolysis of **14**, a mechanism to generate the radical **18** might be involved in the formation of **6** and **17** in the citral solution. In addition, the formation of **14** in the citral solution suggests the possible formation of the corresponding peroxy radical **20**. Therefore, we assumed that the radical **18** might be generated from the peroxy radical **20**, not via hydroperoxide **14**, in the citral solution.

It is generally accepted that alkoxy radicals can be formed directly from peroxy radicals, not via hydroperoxides (20–25). In the case of the formation of **18** from **20**, there seem to be two possible pathways: one is the bimolecular self-coupling of the peroxy radical **20** followed by the decomposition of the resulting tetroxide (20–22) (*path A*), and the other is the addition

Scheme 5. Coupling Reaction between the Phenoxy Radical Derived from EGCg and Peroxy Radicals (ROO•) Followed by the Decomposition of the Coupling Product (37)



Scheme 6. Possible Radical-Scavenging Mechanisms of Phenolic Antioxidants (ArOH) on the Formation of Oxidation Products from Citral



In the present case, as shown in **Scheme 6**, H donation from the catechins to the peroxy radical **20** can afford **14**, which subsequently degrades to **3** and **7** along with **4** under acidic aqueous conditions. On the other hand, coupling reactions between phenoxy radicals derived from the catechins and the peroxy radical **20** followed by the decomposition of the coupling products might produce **7** and **4**.

The observed promoting effects of the catechins seem to be the results of these competing reactions with the peroxy radical **20**. The phenoxy-peroxy coupling reactions that could afford **7** and **4** might be inhibited by the H donation to **20** from the catechins. Therefore, the H donor abilities of the catechins would contribute to the promotion of the formation of **3** and **14** rather than that of **7** and **4**. This might account for the strong promoting effects of the gallated catechins on the formation of **3** and **14**. The importance of the 3-galloyl moiety of the catechins on their H donor abilities has been established in the literature (32–37).

On the other hand, the phenoxy-peroxy coupling reactions might account for the strong promoting effects of the nongallated catechins on the formation of **7** and **4**. This assumption might also be supported by the work of Valcic et al. (31). They concluded that the B-ring rather than the galloyl moiety is the active site for the phenoxy-peroxy coupling reactions of the catechins, because no product resulting from the oxidation of the galloyl moiety was detected when EGCg reacted with peroxy radicals.

Both of the radical scavenging mechanisms of the catechins, that is, H donation and phenoxy-peroxy coupling reactions, against the peroxy radical **20** could contribute to the inhibition of the formation of **6** and **17** according to our proposed mechanism (**Scheme 3**). Further studies including structural elucidations for the resulting oxidation products of the catechins might provide some evidence to support the proposed mechanism for the radical scavenging actions of the catechins in the citral solution.

Quercitrin (quercetin 3-O-rhamnoside) showed strong promoting effects on the formation of **3** and **14**, as much as those of the gallated catechins, yet did not inhibit the formation of **6** and **17** as much as the catechins. This might be accounted for by the lack of a promoting effect of quercitrin on the formation

of **7** and **4**. Because of the electron delocalization due to the 2,3-double bond of the C-ring of the quercetin moiety (38), phenoxy radicals derived from quercitrin might be less reactive to the peroxy radical **20** as compared to those derived from the catechins.

Ascorbic acid also did not inhibit the formation of **6** and **17** as much as the catechins, but the mechanism seems to be different from that of quercitrin. When we conducted the degradation of hydroperoxide **14** (2 mg/L) with added EGC, EGCg, and ascorbic acid (60 mg/L, respectively) under acidic aqueous conditions, only ascorbic acid induced the decomposition of **14** to **6** and **17**. This reduction of **14** by ascorbic acid might be accounted for by the low reduction potential of the ascorbate anion radical (39) compared to those of the catechin phenoxy radicals (40, 41).

In conclusion, this study provided a specific insight into the formation mechanism of the potent off-odorant *p*-methylacetophenone (**6**) from citral under acidic aqueous conditions. Our data suggested that **6** is formed via the *tert*-alkoxy radical **18**, which is most likely derived from the corresponding peroxy radical **20**. Further studies including direct detection of the free radicals by electron spin resonance spectroscopy might provide unambiguous evidence to support our proposed mechanism for the formation of **6** from citral.

ACKNOWLEDGMENT

We thank Susumu Kiyohara, Shuichi Muranishi, and Kenji Kumazawa for helpful discussions and Yuya Sekiguchi for excellent technical assistance.

LITERATURE CITED

- (1) Clark, B. C.; Powell, C. C.; Radford, T. The acid-catalyzed cyclization of citral. *Tetrahedron* **1977**, *33*, 2187–2191.
- (2) McHale, D.; Laurie, W. A.; Baxter, R. L. A reappraisal of the acid-catalyzed cyclization of citral. In *Proceedings of the 7th International Congress of Essential Oils*, Kyoto, 1977; Japan Flavor and Fragrance Manufacturers Association: Tokyo, Japan, 1979; pp 250–253.
- (3) Kimura, K.; Iwata, I.; Nishimura, H. Studies on the deterioration mechanism of lemon flavor. Part I. Relationship between acid-catalyzed cyclization of citral and deterioration of lemon flavor. *Agric. Biol. Chem.* **1982**, *46*, 1387–1389.
- (4) Kimura, K.; Nishimura, H.; Iwata, I.; Mizutani, J. Deterioration mechanism of lemon flavor. 2. Formation mechanism of off-odor substances arising from citral. *J. Agric. Food Chem.* **1983**, *31*, 801–804.
- (5) Peacock, V. E.; Kuneman, D. W. Inhibition of the formation of α ,*p*-dimethylstyrene and *p*-cymen-8-ol in a carbonated citral-containing beverage system. *J. Agric. Food Chem.* **1985**, *33*, 330–335.
- (6) Schieberle, P.; Ehrmeier, H.; Grosch, W. Aroma compounds resulting from the acid catalyzed breakdown from citral. *Z. Lebensm. Unters. Forsch.* **1988**, *187*, 35–39.
- (7) Schieberle, P.; Grosch, W. Identification of potent flavor compounds formed in an aqueous lemon oil/citric acid emulsion. *J. Agric. Food Chem.* **1988**, *36*, 797–800.
- (8) Clark, B. C., Jr.; Chamblee, T. S. Acid-catalyzed reactions of citrus oils and other terpene-containing flavors. In *Off-flavors in Foods and Beverages*; Charalambous, G., Ed.; Elsevier: Amsterdam, The Netherlands, 1992; pp 229–285.
- (9) Ueno, T.; Masuda, H.; Muranishi, S.; Kiyohara, S.; Sekiguchi, Y.; Ho, C.-T. Inhibition of the formation of off-odour compounds from citral in an acidic aqueous solution. In *Flavour Research at the Dawn of the Twenty-first Century*; Le Quéré, J. L., Etiévant, P. X., Eds.; Proceedings of the 10th Weurman Flavour Research Symposium; Lavoisier: Cachan, France, 2003; pp 128–131.

- (10) Creary, X.; Wang, Y.-X. Solvolytic kinetic studies by fluorine-19 NMR. *J. Org. Chem.* **1992**, *57*, 4761–4765.
- (11) Buchanan, M. S.; Connolly, J. D.; Rycroft, D. S. Two new monoterpene hydroperoxides from the liverwort *Jungermannia obovata*. *J. Indian Chem. Soc.* **1998**, *75*, 613–615.
- (12) Kropf, H.; Wischer, D. *p*-Isopropylbenzyl hydroperoxide. Autoxidation of *p*-cymene. *Tetrahedron Lett.* **1969**, 1751–1754.
- (13) Grein, B.; Schmidt, G.; Full, G.; Winterhalter, P.; Schreier, P. 2-Formylmethyl-2-methyl-5-(1-hydroxy-1-methylethyl)tetrahydrofuran: Major volatile product of the water-mediated oxidative decomposition of citral. *Flavour Fragrance J.* **1994**, *9*, 93–98.
- (14) Masuda, H.; Ueno, T.; Muranishi, S.; Irisawa, S.; Ho, C.-T. Inhibition of citral deterioration. In *Free Radicals in Food: Chemistry, Nutrition, and Health Effects*; Morello, M. J., Sahidi, F., Ho, C.-T., Eds.; ACS Symposium Series 807; American Chemical Society: Washington, DC, 2002; pp 176–187.
- (15) Sosnovsky, G.; Rawlinson, D. J. In *Organic Peroxides*; Swern, D., Ed.; Wiley-Interscience: New York, 1971; Vol. 2, pp 186–191.
- (16) Avila, D. V.; Brown, C. E.; Ingold, K. U.; Luszyk, J. Solvent effects on the competitive β -scission and hydrogen atom abstraction reactions of the cumyloxyl radical. Resolution of a long-standing problem. *J. Am. Chem. Soc.* **1993**, *115*, 466–470.
- (17) Fossey, J.; Lefort, D.; Sorba, J. In *Free Radicals in Organic Chemistry*; Wiley: New York, 1995; Chapter 13.
- (18) Goosen, A.; McClelland, C. W.; Rinaldi, F. C. Cyclization of 3-(*p*-methylphenyl)propan-1-ol via its alkoxyl radical and aryl radical cation intermediates. A comparison of regioselectivities. *J. Chem. Soc., Perkin Trans. 2* **1993**, 279–281.
- (19) Citterio, A. Electron-transfer processes by peroxydisulphate: Homolytic benzylation of quinones by alkylarenes and reactions of aromatic radical cations with aromatics. *Gazz. Chim. Ital.* **1980**, *110*, 253–258.
- (20) Blanchard, H. S. A study of the mechanism of cumene autoxidation. Mechanism of the interaction of *t*-peroxy radicals. *J. Am. Chem. Soc.* **1959**, *81*, 4548–4552.
- (21) Bartlett, P. D.; Traylor, T. G. Oxygen-18 tracer studies of alkylperoxy radicals. I. The cumylperoxy radical and chain termination in the autoxidation of cumene. *J. Am. Chem. Soc.* **1963**, *85*, 2407–2410.
- (22) Bartlett, P. D.; Guaraldi, G. Di-*tert*-butyl trioxide and di-*tert*-butyl tetroxide. *J. Am. Chem. Soc.* **1967**, *89*, 4799–4801.
- (23) Mayo, F. R. Free radical autoxidations of hydrocarbons. *Acc. Chem. Res.* **1968**, *1*, 193–201.
- (24) Ingold, K. U. Peroxy radicals. *Acc. Chem. Res.* **1969**, *2*, 1–9.
- (25) Howard, J. A. In *Peroxy Radicals*; Alfassi, Z. B., Ed.; Wiley: New York, 1997; Chapter 10.
- (26) Barton, D. H. R.; Parekh, S. I. A short and efficient method for the preparation of α ,*p*-dimethylstyrene from citral (1). *Synth. Commun.* **1989**, *19*, 3353–3361.
- (27) Andemichael, Y. W.; Wang, K. K. A facile synthesis of 5-methylene-1,3-cyclohexadienes (*o*-isotoluenes) and 1,2,4,6-heptatetraenes. *J. Org. Chem.* **1992**, *57*, 796–798.
- (28) Boozer, C. E.; Hammond, G. S.; Hamilton, C. S.; Sen, J. N. Air oxidation of hydrocarbons. II. The stoichiometry and fate of inhibitors in benzene and chlorobenzene. *J. Am. Chem. Soc.* **1955**, *77*, 3233–3237.
- (29) Liebler, D. C.; Baker, P. F.; Kaysen, K. L. Oxidation of vitamin E: Evidence for competing autoxidation and peroxy radical trapping reactions of the tocopheroxyl radical. *J. Am. Chem. Soc.* **1990**, *112*, 6995–7000.
- (30) Arora, A.; Valcic, S.; Cornejo, S.; Nair, M. G.; Timmermann, B. N.; Liebler, D. C. Reactions of genistein with alkylperoxy radicals. *Chem. Res. Toxicol.* **2000**, *13*, 638–645.
- (31) Valcic, S.; Burr, J. A.; Timmermann, B. N.; Liebler, D. C. Antioxidant Chemistry of Green Tea Catechins. New Oxidation Products of (–)-Epigallocatechin Gallate and (–)-Epigallocatechin from Their Reactions with Peroxy Radicals. *Chem. Res. Toxicol.* **2000**, *13*, 801–810.
- (32) Yoshida, T.; Mori, K.; Hatano, T.; Okumura, T.; Uehara, I.; Komagoe, K.; Fujita, Y.; Okuda, T. Studies on inhibition mechanism of autoxidation by tannins and flavonoids. *Chem. Pharm. Bull.* **1989**, *37*, 1919–1921.
- (33) Salah, N.; Miller, N. J.; Paganga, G.; Tijburg, L.; Bolwell, G. P.; Rice-Evans, C. Polyphenolic flavanols as scavengers of aqueous phase radicals and as chain-breaking antioxidants. *Arch. Biochem. Biophys.* **1995**, *322*, 339–346.
- (34) Guo, Q.; Zhao, B.; Li, M.; Shen, S.; Xin, W. Studies on protective mechanisms of four components of green tea polyphenols against lipid peroxidation in synaptosomes. *Biochim. Biophys. Acta* **1996**, *1304*, 210–222.
- (35) Guo, Q.; Zhao, B.; Shen, S.; Hou, J.; Hu, J.; Xin, W. ESR study on the structure-antioxidant activity relationship of tea catechins and their epimers. *Biochim. Biophys. Acta* **1999**, *1427*, 13–23.
- (36) Nanjo, F.; Goto, K.; Seto, R.; Suzuki, M.; Sakai, M.; Hara, Y. Scavenging effects of tea catechins and their derivatives on 1,1-diphenyl-2-picrylhydrazyl radical. *Free Radical Biol. Med.* **1996**, *21*, 895–902.
- (37) Nanjo, F.; Mori, M.; Goto, K.; Hara, Y. Radical scavenging activity of tea catechins and their related compounds. *Biosci., Biotechnol., Biochem.* **1999**, *63*, 1621–1623.
- (38) Bors, W.; Heller, W.; Michel, C.; Saran, M. Flavonoids as antioxidants: Determination of radical-scavenging efficiencies. *Methods Enzymol.* **1990**, *186*, 343–355.
- (39) Williams, N. H.; Yandell, J. K. Outer-sphere electron-transfer reactions of ascorbate anions. *Aust. J. Chem.* **1982**, *35*, 1133–1144.
- (40) Jovanovic, S. V.; Steenken, S.; Tosic, M.; Marjanovic, B.; Simic, M. G. Flavonoids as antioxidants. *J. Am. Chem. Soc.* **1994**, *116*, 4846–4851.
- (41) Jovanovic, S. V.; Hara, Y.; Steenken, S.; Simic, M. G. Antioxidant potential of gallic catechins. A pulse radiolysis and laser photolysis study. *J. Am. Chem. Soc.* **1995**, *117*, 9881–9888.

Received for review December 26, 2003. Revised manuscript received June 16, 2004. Accepted June 20, 2004.

JF035517J